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ΕΡΕΥΝΗΤΙΚΗ ΕΡΓΑΣΙΑ

Does celecoxib, a COX-2 inhibitor, spare the traditional nonsteroidal anti-inflammatory-mediated adverse events?

OBJECTIVE To investigate the gastrointestinal (GI) and reproductive risk associated with chronic usage of celecoxib in male albino rats. **METHOD** Thirty male albino rats weighing 200–230 g were randomly divided into three equal groups of 10 rats each. In group 1, the control group, rats received no drug. In group 2, rats received celecoxib (50 mg/kg/day, orally) for 30 successive days. In group 3, rats received celecoxib (50 mg/kg/day, orally), and royal jelly (300 mg/kg/day, orally) for 30 successive days. Gastric ulcer scoring and semen analysis were employed to elucidate the celecoxib adverse effects on gastric and testicular tissue functions. **RESULTS** Celecoxib produced no gastric ulcer in both groups 2 and 3. Celecoxib has no appreciable effect on steroidogenesis (testosterone biosynthesis in Leydig cells of the testis) or spermatogenesis (sperms production from germ cells in the seminiferous tubules of the testis), as indicated by normal sperm count, motility, abnormalities, and normal serum testosterone levels. Royal jelly co-administration did not cause any significant change in serum testosterone level, sperm count, and sperm motility percentage, but caused a significant decline in sperm abnormalities level. **CONCLUSIONS** Upper GI tolerability and reproductive safety were proved in the chronic use of celecoxib in male albino rats, with ameliorative effects of royal jelly against celecoxib-induced oxidative and apoptotic stress.

The Food and Drug Administration (FDA) has given the drug family known as non-steroidal anti-inflammatory drugs (NSAIDs) approval for use as analgesic, antipyretic, and anti-inflammatory medications.¹ NSAIDs are often categorized into non-selective cyclooxygenase (COX) inhibitors (such as diclofenac, ibuprofen, indomethacin, ketoprofen, etc.) and selective COX-2 inhibitors (such as celecoxib), depending on their chemical structure and selectivity.²

The epidemiological and clinical literature has extensively covered the gastrointestinal (GI) adverse effects linked to the use of non-steroidal anti-inflammatory drugs. The COX enzyme is inhibited by NSAIDs, which are thought to harm the gastro-duodenum by inhibiting the COX-1 isoform. Celecoxib, a COX-2 selective inhibitor, has been demonstrated to be related to decreased gastroduodenal ulcer rates when compared to standard NSAIDs.³ With

greater GI tolerance than traditional NSAIDs, celecoxib may maintain similar analgesic and anti-inflammatory activity while being a better alternative for patients at risk of GI bleeding.⁴ This is evidenced by lower incidence rates of gastric, duodenal, and gastroduodenal ulcers.³

It has been established that COX-2 is a key player in the negative autoregulation of steroidogenesis, presumably through the synthesis of PGE2 and PGF2.⁵ Due to COX-2 suppression in the adult rat testis, celecoxib has been demonstrated in several studies to have direct or indirect effects on spermatogenesis.^{6,7} Celecoxib taken at high doses has been shown to decrease the size and number of Leydig cells, which lowers testosterone levels.⁸ As a result, prolonged high celecoxib doses are linked to male infertility.⁹ Conversely, according to other studies, celecoxib has no discernible impact on steroidogenesis or

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Με τη χρήση σελεκοξίμπης, ειδικού
αναστολέα της COX-2,
αποφεύγονται οι ανεπιθύμητες
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συνήθως από τα μη
συνταγογραφούμενα
αντιφλεγμονώδη φάρμακα;

Περίληψη στο τέλος του άρθρου

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spermatogenesis, as evidenced by normal testicular weight, testicular morphology, or serum testosterone levels,¹⁰ as well as normal sperm concentration, motility, and DNA fragmentation.^{11,12} Men with normal spermatogenesis may benefit from being advised to take chronic celecoxib, but subfertile or infertile men should not.¹³

Royal jelly (RJ) is a nutrient-rich source of bioactive compounds that are essential to many biological processes. It has been determined that proteins, peptides, lipids, phenolics, and flavonoids are the main bioactive compounds responsible for the variety of pharmacological properties of RJ.¹⁴ The use of RJ in the treatment of eye disorders, GI tract diseases, neurological disorders, fertility disorders, and wound healing activity was advised by its potential health benefits, such as antimicrobial, antioxidant, anti-inflammatory, anticancer, antihyperlipidemic, cardioprotective, and hepatorenal-protective properties.¹⁵

Therefore, we studied the safety issue of celecoxib towards gastric ulcers and male fertility in male albino rats with special concern for the potential effect of RJ administration.

MATERIAL AND METHOD

Experimental design

Thirty, apparently healthy, male albino rats weighing 200–230 g were employed in this study. They were obtained from the laboratory animal farm, Faculty of Veterinary Medicine, Zagazig University. They were contained in clean cages with wire-bottomed galvanised metal walls. Throughout the experiment, they were given a balanced diet and unlimited access to fresh tap water. Before being enrolled in the study, they had a 7-day acclimatisation period. Rats were randomly divided into three equal groups; each of 10 rats: (a) Group 1 (control group): Rats in this group received no drug. Rats of this group served as the control group. (b) Group 2 (celecoxib group): Rats in this group received celecoxib (50 mg/kg/day, orally)¹⁶ for 30 successive days, then rats were sacrificed and samples were collected. (c) Group 3 (celecoxib plus RJ group): Rats in this group received celecoxib (50 mg/kg/day, orally),¹⁶ and RJ (300 mg/kg/day, orally)¹⁷ for 30 successive days, then rats were sacrificed and samples were collected.

Samples collection and preservation

Blood samples

Blood samples were collected using a 3 mL syringe directly from the ventricular puncture of rats into centrifuge tubes and left to clot for 15 min at room temperature then centrifuged at 3,000 rpm for 10 minutes to allow serum separation, which was then aspirated into cryovials and stored at -20 °C for serum biochemical assays.

Tissue specimens

The liver, kidneys, heart, stomach, and testes tissues of each animal were dissected and collected as follows: (a) Gastric ulcer scoring: The stomach was quickly washed in an ice-cold saline solution (0.9% NaCl), containing 0.16 mg/mL heparin to remove red blood cells and a dissecting microscope with a square-grid eyepiece was used to quantify rats' stomach ulceration levels.¹⁸

Semen collection: One testis' cauda epididymis was quickly taken out and put in a sterile Petri plate with 2 mL of sodium chloride 0.9% solution at 37 °C. The epididymal contents were then extracted into the solution to create a suspension using a sterile scissor for semen analysis.¹⁹

Biological analysis

About 30 mg of the rat tissue samples were immediately collected, transferred in liquid nitrogen, and kept at -80 °C for total RNA extraction used for the determination of apoptotic and anti-apoptotic mRNA expression levels using RT-PCR,²⁰ for detection of apoptotic genes (*Bax*) and anti-apoptotic gene (*Bcl-2*).²¹

Biochemical measurements in sera

- Fertility tests. Measurement of serum testosterone concentration:* Quantitative estimation of testosterone (TH) concentration in serum was carried out using a commercially available Abnova® ELISA kit, according to the method of Granoff and Abraham.²²
- Oxidant/antioxidant status:* Commercially available kits supplied from Oxi Select™, USA, were used for biochemical quantitative estimation of serum malondialdehyde (MDA), and superoxide dismutase (SOD), according to the method of Wang et al.²³

Quantification of gastric ulceration

Rats' stomach ulceration levels were measured using the Szabo and Hollander technique.¹⁸ In brief, stomachs that had been cleaned were pinned to a corkboard, and ulcers were graded on a 0–5 scale (depicting the severity of vascular congestions and lesions/hemorrhagic erosions). The ulcers were scored as follows: Almost normal mucosa: 0; vascular congestions: 1; one or two lesions: 2; severe lesions: 3; very severe lesions: 4; perforation: 5.

Mucosal damage was indicated as a percentage of the glandular stomach's overall surface area, which was calculated in square millimeters. Each rat's mean ulcer score was expressed as ulcer index (UI) and the percentage of inhibition against ulceration was determined using the expressions:

$$UI = [\text{Ulcerated area} / \text{total stomach area}] \times 100$$

$$\% \text{Ulcer inhibition} = [UI \text{ in control} - UI \text{ in test}] \times 100 / UI \text{ in control}$$

Semen analysis

The rat epididymal contents were extracted into 2 mL of sodium

chloride 0.9% solution in a sterilized Petri dish at 37 °C making semen suspension. Under a 40× high-power light microscope, sperm motility was analyzed. The sperm count was conducted at 100× magnification under a light microscope. The following formula was used to determine how many sperms were present in 1 mL of the fluid:

$$\text{Mean number of sperm in each chamber} \times 10,000 \times \text{dilution factor}$$

A microscope was used to assess morphological changes at 400× magnification. To measure the proportion of abnormal spermatozoa, about 100 spermatozoa were randomly observed in several fields while submerged in oil under an oil immersion lens. The following sperm traits were utilized to categorize them as abnormal: bent neck, bent tail, detached head, coiled tail, double head, amorphous head, curved tail, fused, looped tail, looped neck, detached tail, and hookless sperms.²⁴

Apoptotic and anti-apoptotic genes expression

TOPreal™ qPCR 2X PreMIX (SYBR Green with low ROX) (Cat. # P725, Enzynomics, Korea) was used for quantitative estimation of apoptotic genes (*Bax*) and anti-apoptotic gene (*Bcl-2*) based on the manufacturer's protocol.

Gene expressions were measured using the below formula and Ct (2- $\Delta\Delta$ Ct) (fold change) method Eleawa et al.²⁰

$$\Delta\Delta Ct = (Ct_{\text{target}} - Ct_{\text{reference}})_{\text{test sample}} - (Ct_{\text{target}} - Ct_{\text{reference}})_{\text{control sample}}$$

Finally, considering the primer efficiency value of approximately 2, the gene expression level was determined as 1- $\Delta\Delta$ Ct.

Statistical analysis

The obtained data were analyzed statistically using the Statistical Package for Social Sciences (SPSS) program (version 26.0; SPSS Inc, IL, USA) for Microsoft Windows®. Results were expressed as the mean±standard deviation. One-way analysis of variance (ANOVA) was used to compare the means of the multiple groups in the study. Tukey HSD *post hoc* analysis was used for in-between

group comparisons, where a p-value of less than 0.05 is considered to be significant.

RESULTS

Quantification of gastric ulceration

No gastric ulceration was detected in any of the study groups (fig. 1).

Fertility profile

Chronic celecoxib administration has no appreciable effect on testosterone hormone levels or spermatogenesis (sperms production from germ cells in the seminiferous tubules of the testis) as indicated by normal serum testosterone levels, normal sperm count, motility, and abnormalities. Royal jelly co-administration did not cause any significant change in serum testosterone level, sperm count, and motility percentage, but caused a significant decline in sperm abnormalities level (tab. 1).

Oxidant/anti-oxidant status

Chronic celecoxib administration impaired oxidant/antioxidant status as indicated by significant elevation ($p < 0.05$) in serum MDA level, and significant decline ($p < 0.05$) in serum SOD level. Royal jelly co-administration has a positive effect on oxidative status as indicated by a significant decline ($p < 0.05$) in serum MDA level, and significant elevation ($p < 0.05$) in serum SOD level (tab. 2).

Apoptotic and anti-apoptotic genes expression

Chronic celecoxib administration significantly upregulated ($p < 0.05$) apoptotic gene (*Bax/Gapdh* %) expression



Figure 1. Effects of celecoxib (50 mg/kg/day, orally), royal jelly co-administration (300 mg/kg/day, orally), on gastric mucosa: No gastric ulcers. **Group 1 (G1)** received no drug (control). **Group 2 (G2)** received celecoxib (50 mg/kg/day, orally) for 30 successive days. **Group 3 (G3)** received celecoxib (50 mg/kg/day, orally) and royal jelly (300 mg/kg/day, orally) for 30 successive days.

and significantly downregulated ($p < 0.05$) anti-apoptotic gene (*Bcl-2/Gapdh* %) expression levels in the stomach, and testis. Royal jelly co-administration significantly down-regulated ($p < 0.05$) apoptotic gene (*Bax/Gapdh* %) expression and significantly upregulated ($p < 0.05$) anti-apoptotic

gene (*Bcl-2/Gapdh* %) expression levels in the stomach, and testis (tab. 3).

DISCUSSION

Celecoxib is a NSAID used to treat mild to moderate pain and help relieve symptoms of various forms of arthritis and the management of acute or chronic pain due to its favorable GI toxicity profile.²⁵ Data indicate that celecoxib has no appreciable effect on steroidogenesis or spermatogenesis, at least in the short term, but may have implications for men with marginal fertility taking celecoxib for extended periods, especially in case of subfertility.¹⁰ Celecoxib treatment mitigates oxidative stress,²⁶ but induces apoptosis.²⁷ Fortunately, RJ can ameliorate NSAIDs side effects.¹⁷

In our study, celecoxib administration (50 mg/kg/day, orally) for 30 successive days did not cause gastric ulcers in male albino rats. Celecoxib, a COX-2 inhibitor, does not block the constitutive COX-1 enzyme,²⁸ which is responsible for GI tract prostaglandins expression, that ensures gastric mucosal safety,²⁹ whereas it blocks the inducible COX-2 expression that occurs in case of tissue injury.³⁰

Two different approaches were pursued in the search of GI sparing NSAIDs: (a) Classical COX hypothesis, where only COX-1 is responsible for gastric stomach integrity, and (b) modified COX hypothesis, where both COX-1 and COX-2 are involved in the protection of gastric mucosa.

In the classical COX hypothesis, COX-1-derived prostaglandins are considered to exert housekeeping functions in the gastric mucosa essential for normal gastric physiology. Inhibition of prostaglandin formation leads to a decrease in mucus and bicarbonate secretion, reduces mucosal blood flow, and causes vascular injury, leukocyte accumulation, and reduced cell turnover, all factors that contribute to the genesis of mucosal damage.³¹ Selective COX-2 inhibitors do

Table 1. Effects of celecoxib (50 mg/kg/day, orally), and royal jelly (300 mg/kg/day, orally) on fertility profile in male albino rats. Data are expressed as mean \pm SD, n=10 per group.

Groups	Fertility profile			
	Testosterone (ng/mL)	Sperm motility (%)	Sperm count ($\times 10^6$)	Sperm abnormality (%)
1 (n=10)	3.30 \pm 1.85*	82.00 \pm 5.00*	93.33 \pm 37.17*	14.33 \pm 5.13***
2 (n=10)	2.60 \pm 0.65*	80.63 \pm 2.89*	89.00 \pm 7.81*	18.33 \pm 2.31*
3 (n=10)	2.90 \pm 0.40*	81.00 \pm 5.00*	90.33 \pm 4.16*	14.67 \pm 6.51***

Group 1 received no drug (control)

Group 2 received celecoxib (50 mg/kg/day, orally) for 30 successive days

Group 3 received celecoxib (50 mg/kg/day, orally) and royal jelly (300 mg/kg/day, orally) for 30 successive days

Means carrying different superscripts in the same column are significant at $p < 0.05$
SD: Standard deviation

Table 2. Effects of celecoxib (50 mg/kg/day, orally), and royal jelly (300 mg/kg/day, orally) on serum oxidative status profile in male albino rats. Data are expressed as mean \pm SD, n=10 per group.

Groups	MDA (nmol/mL)	SOD (U/mL)
1 (n=10)	5.86 \pm 0.66***	5.40 \pm 0.62*
2 (n=10)	28.10 \pm 3.48*	0.89 \pm 0.68***
3 (n=10)	16.03 \pm 2.51**	3.20 \pm 0.61**

Group 1 received no drug (control)

Group 2 received celecoxib (50 mg/kg/day, orally) for 30 successive days

Group 3 received celecoxib (50 mg/kg/day, orally) and royal jelly (300 mg/kg/day, orally) for 30 successive days

Means carrying different superscripts in the same column are significant at $p < 0.05$
SD: Standard deviation, MDA: Malondialdehyde, SOD: Superoxide dismutase

Table 3. Effects of celecoxib (50 mg/kg/day, orally), and royal jelly (300 mg/kg/day, orally) on apoptotic and anti-apoptotic genes expression in male albino rats. Data are expressed as mean \pm SD, n=10 per group.

Groups	Apoptotic gene (<i>Bax/Gapdh</i> %)		Anti-apoptotic gene (<i>Bcl-2/Gapdh</i> %)	
	Stomach	Testis	Stomach	Testis
1 (n=10)	1.00 \pm 0.00***	1.00 \pm 0.00***	1.00 \pm 0.02*	1.00 \pm 0.00*
2 (n=10)	5.74 \pm 1.03*	6.16 \pm 1.3*	0.19 \pm 0.06***	0.30 \pm 0.03***
3 (n=10)	1.90 \pm 0.19***	2.35 \pm 0.18***	0.57 \pm 0.00***	0.81 \pm 0.12***

Group 1 received no drug (control)

Group 2 received celecoxib (50 mg/kg/day, orally) for 30 successive days

Group 3 received celecoxib (50 mg/kg/day, orally) and royal jelly (300 mg/kg/day, orally) for 30 successive days

Means carrying different superscripts in the same column are significant at $p < 0.05$
SD: Standard deviation

not affect COX-1 in the GI tract mucosa. They appear to be well tolerated by experimental animals and humans following acute and chronic (three or more months) administration. Thus, selective COX-2 inhibitors have been developed as GI-sparing anti-inflammatory drugs.^{32,33}

The modern, modified COX hypothesis, has accused its predecessor of being oversimplistic.³⁴ It assumed that COX-1 and low levels of COX-2 contribute to gastric mucosal integrity. COX-1-derived prostaglandins regulate the mucosal blood flow and epithelial secretion of mucus and bicarbonate, while COX-2-derived prostaglandins affect epithelial proliferation and endothelial-leukocyte adherence,³⁵ which plays an important role in the healing of gastric ulcers.³⁴ Recent studies in rats have shown that neither selective COX-1 inhibition nor selective COX-2 inhibition is ulcerogenic individually.³⁶ Only combined inhibition of both COX-1 and COX-2 induces severe lesions in the stomach and small intestine, suggesting that reduction of mucosal blood flow (COX-1 inhibitors) and increase in leukocyte adhesion (COX-2 inhibitors) have to occur simultaneously to interfere with gastric mucosal defense.^{37,38} These findings are confirmed by experiments using COX-1- or COX-2-deficient mice. Neither COX-1-knockout mice,³⁹ nor COX-2-deficient mice showed spontaneous GI lesions or even gastric pathology.⁴⁰

Another advantage of celecoxib has been shown not to interfere with platelet function because it does not inhibit platelet thromboxane A₂ formation, which could be a contributory factor in the reduced incidence of GI hemorrhage.⁴¹ Another not widely recognized advantage is its lack of interference with mitochondrial oxidative phosphorylation responsible for normal mucosal prostanoid levels important to gastric mucosa integrity.⁴²

In addition to prostaglandins, other mediator systems, including nitric oxide (NO) have the potential to maintain gastric mucosal integrity in case of inhibition of prostaglandin synthesis⁴³ via inhibition of leukocyte adhesion to the endothelium, and healing of NSAID-induced damage.⁴⁴ Celecoxib beneficially lowers the expression of COX-2, hence reducing oxidative stress and increasing the bioavailability of NO.⁴⁵ Neither the COX-1 inhibitor (SC-560) nor COX-2 inhibitors (NS-398 or DFU) individually could cause stomach ulcers in the Ehrlich et al study. However, severe and dose-dependent injury developed when simultaneously the NO synthase was suppressed. Gastric mucosal damage induced by COX-2 inhibitors was only found after pre-treatment with a NO synthase inhibitor (L-NAME).⁴⁶ Celecoxib produced no gastric ulcer as it has been developed with several hundred-fold higher selectivity for COX-2 with

no effect on COX-1 prostaglandin production responsible for gastric mucosa integrity.⁴⁷

In accordance with our findings, studies have confirmed a better GI profile of celecoxib.^{48,49} The incidence of GI ulcer complications associated with celecoxib was 8-fold lower than with nonspecific NSAIDs.⁵⁰ In patients at high risk of GI events who require concomitant aspirin and NSAID, celecoxib plus proton-pump inhibitor is the preferred treatment to reduce the risk of recurrent upper GI bleeding.⁵¹ Treatment of gastric cancer with celecoxib is not associated with an increased risk of gastroduodenal ulcer.^{52,53} Gastrointestinal tolerability of celecoxib in elderly patients aged 65 years or older with osteoarthritis, rheumatoid arthritis, or ankylosing spondylitis was higher in celecoxib than in conventional nonselective NSAIDs.⁵⁴ Celecoxib was found to be more effective in post-endodontic pain with minimal gastric symptoms, when compared to other NSAIDs.⁵⁵

Without a doubt, RJ has a beneficial effect in preventing stomach ulcers. It inhibits inducible nitric oxide (iNOS) and nuclear factor kappa beta (Nf- κ) activity in the gastric mucosa and prevents epithelial cell apoptosis. It also significantly decreases the activity of pro-inflammatory cytokines, such as tumor necrosis factor- α (TNF- α) and interleukin-1 beta (IL-1 β) levels. In addition, a decrease in the myeloperoxidase (MPO) level with favorable tissue oxidative status indicated that RJ prevented tissue damage.⁵⁶

In our study, celecoxib administration (50 mg/kg/day, orally) for 30 successive days was claimed to cause a non-significant decline in serum testosterone hormone level, sperm motility, and sperm count in male albino rats; in addition to non-significant elevation in sperm abnormalities percentage. An ameliorative effect of RJ against celecoxib-induced infertility.

In accordance with our findings, celecoxib, as a COX-2 inhibitor, has not uncovered obvious effects on normal fertility or testicular function.¹⁰ To comprehend this hypothesis, we must be aware that both COX isoforms, referred to as COX-1 and COX-2, are expressed constitutively in male rat testes,^{57,58} knowing that COX-2 is predominant.⁵⁹ The localization and expression levels of COX-2 significantly differ between the species. In rat testis, COX-2 has been found to localize in germ and Leydig cells, while in mice it is restricted to interstitial cells of Leydig cells.⁶⁰ In humans, discordant data are reported. While Frungieri et al confirmed the absence of COX-2 in male testes indicating impaired spermatogenesis if present,⁶¹ Perrotta et al confirmed that both COX-1 and COX-2 are expressed and present in humans with normal spermatogenesis and COX-2 levels

are markedly increased in inflamed tissues as in varicocele and diabetic human donor's sperm.⁶²

Undoubtedly, COX-2 possesses an important role in male reproduction activity via PGs production, which has important roles in both physiological and pathological processes in the testes.⁶³ Endogenous production of PGE₂ and PGF_{2α} by the testis can be attributed to COX-2 rather than COX-1.¹¹ COX-2/PGs are expressed in the two key somatic cell types in the testis, Leydig and Sertoli cells, under the regulation of hormonal input (FSH, prolactin, and testosterone), as well as by IL1β for the production of steroidogenesis in Leydig cells and glucose uptake in Sertoli cells. Hence, such COX-2/PG system acts as a local modulator of testicular activity and consequently may regulate spermatogenic efficiency.^{12,64}

Regarding celecoxib, studies indicate both positive and negative effects of celecoxib in the preservation of male fertility. Chronic celecoxib would be of little consequence for males with optimal fertility. Oral administration of celecoxib, for 5 weeks, increases testicular interstitial fluid (IF) formation in the testis as a result of an increase in either testicular blood flow, capillary pressure, or vascular permeability, consistent with inhibition of rat testicular vasculature PGs production, but has no appreciable effect on steroidogenesis (steroid hormones, including testosterone, biosynthesis from cholesterol in Leydig cells of the testis) or spermatogenesis (sperms production from germ cells in the seminiferous tubules of the testis). Normal testis weight, testis morphology, serum testosterone levels, and normal sperm concentration, motility, and DNA fragmentation are confirmed.^{10,13,65} In addition, celecoxib does not appear to alter the ability of the testis to mount an inflammatory response but opposes the deleterious effects of inflammation on IF formation and testosterone production, so it has the potential to ameliorate testicular damage caused by systemic or local inflammation,¹⁰ and some toxicants in different pathological conditions in conjunction with serum and testis inflammatory cytokines, oxidative stress, or genetic alterations.⁶⁶ In this context, celecoxib offers protection against reproductive abnormalities induced by atrazine in male rats,⁶⁷ and against varicocele-induced damage at testicular and sperm levels in male rats.^{68,69}

In contrast, celecoxib's negative impact on fertility is frequently associated with males with pre-existing subfertility or infertility. Increased testicular IF accumulation in the testis with increased capillary pressure causes inhibition of rat testicular vasculature PGs production and inhibition of T-type Ca²⁺ channels in spermatogenic cells which affects

the Ca²⁺, pH, and membrane potential homeostasis of sperm cells.^{10,70}

The effect of RJ on sperm parameters and testosterone levels was studied extensively. The results were consistent with what our study found. Royal jelly (200 mg/kg, 400 mg/kg, and 800 mg/kg daily for 4 weeks, orally) in male Sprague-Dawley rats did not cause any significant change in sperm count because of all doses. No significant change in serum testosterone level or sperm deformity rate except for the high-dose group. High-dose RJ oral administration for 4 weeks adversely affected the reproductive system, in the form of an increase in the levels of estrogen, LH, FSH, and a decrease in testosterone level, which returned to normal levels after 14 days of RJ cessation.⁷¹ Royal jelly (100 mg/kg daily for 48 days, orally) in male Wistar rats caused no change in sperm count, viability, motility, or testosterone production.⁷² Royal jelly (100 mg/kg daily for 30 days, orally) in male mice caused a non-significant increase in sperm count, motility, and plasma testosterone levels.⁷³ Testosterone levels were non-significantly improved in men who received short-term RJ supplement (1,000 mg/daily/15 days),⁷⁴ long-term RJ supplement (100 mg daily for two months),⁷⁵ and increasing long-term RJ supplement (25 mg/kg, 50 mg/kg, and 100 mg daily for three months).⁷⁷ Interestingly, no significant difference in sperm count before and after consumption of RJ in all of the above investigations is reported.⁷⁴⁻⁷⁶ Fortunately, RJ (100 mg/kg orally) significantly reduced the sperm abnormalities percentage in optimal⁷⁷ and high ambient temperature⁷⁸ conditions.

In the present study, celecoxib administration (50 mg/kg/day, orally) for 30 successive days was claimed to cause oxidative stress (OS) in male albino rats as manifested by significant elevation of serum malondialdehyde (MDA) and significant decline in superoxide dismutase (SOD) levels, with ameliorative effects of RJ against celecoxib-induced oxidative stress.

OS is a shift in the balance between the production rate of oxidants and their elimination via the antioxidant defense system.⁷⁹ OS-mediated molecules are an array of metabolites derived from molecular reactive oxygen species (ROS) and reactive nitrogen species (RNS).⁸⁰ Prime sites of ROS production in living organisms are the mitochondria, peroxisomes, and endoplasmic reticulum. ROS are produced during the mitochondrial electron transport chain, through the oxidative phosphorylation pathway for ATP production.⁸¹ ROS are also produced in phagocytes, auto-oxidation reactions, and subsequent to antioxidant enzymes (e.g., xanthine oxidase).⁸² RNS is derived from nitric oxide (·NO) metabolism.⁸³ ROS/RNS are produced from either endogenous or exogenous sources. Immune system activation, inflammation, mental

stress, excessive exercise, cancer, infection, ischemia, and aging represent endogenous stressors, while air pollution, water pollution, alcohol, cigarette smoke, heavy metals, pharmaceutical agents, and ultraviolet radiation act as exogenous sources of OS.⁸⁴

Antioxidants are the primary line of the body's defense system against oxidative stress. In physiological conditions, antioxidants inhibit the oxidation reaction of molecules that can produce free radicals.⁸⁵ Several molecules play a role in such defense, either internally synthesized (endogenous), or externally supplied through food (exogenous) antioxidants. Exogenous antioxidants (RJ for instance) act as direct ROS scavengers and increase the antioxidant enzyme activities.⁸⁶ Endogenous antioxidant defenses involve a network of antioxidant enzymatic and non-enzymatic molecules that are usually disseminated within the cytoplasm and various cell organelles.⁸⁷ Primary antioxidant enzymes, such as superoxide dismutase (SOD), catalase, and several peroxidases, catalyze a complex cascade of reactions to transform ROS into more stable molecules, such as water and O₂. MDA is the end-product of polyunsaturated fatty acids oxidation in cellular membranes; thus, it acts as a dependable marker of oxidative stress.⁸⁸ Non-enzymatic antioxidants include vitamins (A, C, E, and K), enzyme cofactors (Q10), and minerals (Zn, Mn, Cu, Se, etc.).⁸⁹

Following our findings, previous studies indicated an alteration in the oxidant/antioxidant status of rats treated with celecoxib.⁹⁰⁻⁹³ The excessive generation of free radicals is considered one of the mechanisms by which celecoxib induces toxicity. Free radicals trigger the development of many diseases and cause harmful effects that cause the peroxidation of biomembranes and DNA, which is the reason for tissue destruction.⁹⁴

Previous studies confirmed the concept of the beneficial and antioxidant effects of RJ.⁹⁵⁻⁹⁷ Administration of RJ *in vitro* inhibited the production of pro-inflammatory cytokines, such as TNF- α and IL-1, IL-6 in a dose-dependent manner.⁹⁸ Other *in vivo* and *in vitro* studies also proposed anti-inflammatory and anti-oxidant properties of RJ.⁹⁹ The antioxidant activity of RJ can be attributed to the free radical scavengers; they can inhibit lipid peroxidation¹⁰⁰ via scavenging hydroxyl radicals,¹⁰¹ and they attributed this activity to three dipeptides containing tyrosine residues. Additionally, RJ inhibits the enzymes that elevate the peroxidation of endogenous lipids, as well as cytochrome P450 expression, which is one of the cellular sources of oxygen radicals.¹⁰²

In the present study, celecoxib administration (50 mg/kg/day, orally), for 30 successive days was claimed to induce

an apoptotic effect in male albino rats as manifested by a significant elevation of serum apoptotic gene (*Bax*) and a significant decline in serum anti-apoptotic gene (*Bcl-2*) in stomach, and testis, with ameliorative effects of RJ against the celecoxib-induced apoptotic effect.

Apoptosis is programmed cell death, which involves the genetically determined elimination of cells. Since too little or too much cell death can result in pathology, such as autoimmune illnesses, neurodegeneration, or cancer, it is obvious that apoptosis needs to be strictly gene-regulated.¹⁰³ Several factors contributed to the apoptotic mechanism, among them, two main families of proteins including caspases and the *Bcl-2* family. Caspases, death-driving cysteine proteases, play an effective role in the apoptotic process. Activation of caspases ensures that the cellular components are degraded in a controlled manner, carrying out cell death with minimal effect on surrounding tissues.¹⁰⁴ Whereas, *Bcl-2*, consists of anti-apoptotic and pro-apoptotic members. The anti-apoptotic members of this family, such as *Bcl-2* and *Bcl-XL*, prevent apoptosis either by sequestering caspases or by preventing the release of mitochondrial apoptogenic factors, such as cytochrome c and apoptosis-inducing factor (AIF) into the cytoplasm, which are responsible for caspase activation. In contrast, pro-apoptotic members of this family, such as *Bax* and *Bak*, trigger the release of caspases by inducing the release of mitochondrial apoptogenic factors into the cytoplasm leading to caspase activation.¹⁰⁵

Apoptosis is an essential physiological process for the selective elimination of individual cells without destruction or damage to the whole organ.¹⁰⁶ The morphological features of apoptosis include cell shrinkage and pyknosis, cytoplasmic and nuclear condensation, chromatin cleavage, apoptotic bodies formation, an intact plasma membrane, and exposure of surface molecules to phagocytosis.¹⁰⁷ In contrast, necrosis has been characterized as passive, accidental cell death that is triggered by external factors or disease with the uncontrolled release of inflammatory cells.^{108,109} The main morphological changes associated with necrosis include cell swelling, cytoplasmic vacuole formation, distended endoplasmic reticulum, cytoplasmic bleb formation, condensed, swollen, or ruptured mitochondria, disrupted organelle membranes, swollen and ruptured lysosomes, and ultimately disruption of the cell membrane.¹¹⁰ While necrosis is always pathological to an organ,¹⁰³ apoptosis is a normal and physiological process to get rid of certain cells that have been damaged beyond repair. It is almost always normal and advantageous. Additionally, apoptosis happens as a protective process, such as in immunological responses or when

diseases or toxic chemicals destroy cells. Thus, it keeps normal tissue function.¹¹¹

Following our findings, previous studies have confirmed celecoxib-induced apoptosis. Celecoxib causes apoptosis by causing the loss of the mitochondrial transmembrane potential, the release of cytochrome c and AIF, and the activation of caspase-9 and caspase-3. Additionally, the anti-apoptotic protein Bcl-2 was reduced in abundance whereas the pro-apoptotic protein Bax was enhanced by celecoxib. The data showed that mitochondria-dependent signaling, not PPAR/NF- κ B signaling, was the mechanism through which celecoxib triggered apoptosis in mouse liver cancer cells.²⁷ In another study, celecoxib induced apoptosis in 5-fluorouracil-resistant gastric cancer cells through protein kinase B (PKB) inhibition,¹¹² which is a key component of the phosphatidylinositol-3 kinase (PI3K) intracellular pathway that exerts a pivotal role in regulating cell proliferation, survival, and metabolism.¹¹³

Celecoxib induced apoptosis in glioblastoma tumor cells, the primary malignant tumor of the brain, via suppressing CIP2A/PP2A/Akt signaling axis.¹¹⁴

Several previous studies confirmed the concept of the beneficial and anti-apoptotic effect of RJ against cisplatin-induced hepatorenal toxicity,¹¹⁵ nicotine-induced testicular injury in mice,¹¹⁶ doxorubicin-induced nephrotoxicity in male albino rats,¹¹⁷ and hydroxyurea-induced hepatic injury in rats.¹¹⁸ Moreover, RJ decreases the expression of the apoptotic gene (*MMP-9*) responsible for bladder cancer in humans.

The limitation of our study was the relatively small final number of experimental animals in each group.

In conclusion, upper GI tolerability and reproductive safety were proved in the chronic use of celecoxib in male albino rats, with ameliorative effects of RJ against celecoxib-induced oxidative and apoptotic stress.

ΠΕΡΙΛΗΨΗ

Με τη χρήση σελεκοξίμπης, ειδικού αναστολέα της COX-2, αποφεύγονται οι ανεπιθύμητες ενέργειες που προκαλούνται συνήθως από τα μη συνταγογραφούμενα αντιφλεγμονώδη φάρμακα;

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ΣΚΟΠΟΣ Η διερεύνηση του γαστρεντερικού και του αναπαραγωγικού κινδύνου που σχετίζεται με τη χρόνια χρήση της σελεκοξίμπης (celecoxib) σε άρρνες αλμπίνο (albino) αρουραίους. **ΥΛΙΚΟ-ΜΕΘΟΔΟΣ** Τριάντα αρσενικοί αλμπίνο αρουραίοι, βάρους 200–230 g, χωρίστηκαν τυχαία σε 3 ίσες ομάδες από 10 η κάθε μία: Ομάδα 1 (ομάδα ελέγχου): αρουραίοι που δεν έλαβαν φάρμακο, Ομάδα 2: αρουραίοι που έλαβαν σελεκοξίμπη (50 mg/kg/ημέρα, από το στόμα) για 30 διαδοχικές ημέρες και Ομάδα 3: αρουραίοι που έλαβαν σελεκοξίμπη (50 mg/kg/ημέρα, από το στόμα) και βασιλικό πολτό (300 mg/kg/ημέρα, από το στόμα) για 30 διαδοχικές ημέρες. Η βαθμολογία του γαστρικού έλκους και η ανάλυση σπέρματος χρησιμοποιήθηκαν για τη διερεύνηση των ανεπιθύμητων ενεργειών της σελεκοξίμπης στις λειτουργίες του γαστρικού ιστού και των όρχεων. **ΑΠΟΤΕΛΕΣΜΑΤΑ** Η σελεκοξίμπη δεν παρήγαγε γαστρικό έλκος και στις δύο ομάδες 2 και 3. Η σελεκοξίμπη δεν είχε αξιόλογη επίδραση στη στεροειδογένεση (βιοσύνθεση τεστοστερόνης στα κύτταρα Leydig του όρχεος) ή στη σπερματογένεση (παραγωγή σπέρματος από γεννητικά κύτταρα στα σπερματοζωάρια των όρχεων), όπως υποδείχθηκε από τον φυσιολογικό αριθμό σπερματοζωαρίων, την κινητικότητα, τις ανωμαλίες και τα φυσιολογικά επίπεδα τεστοστερόνης ορού. Η συγχορήγηση βασιλικού πολτού δεν συνοδεύτηκε από κάποια σημαντική αλλαγή στο επίπεδο τεστοστερόνης στον ορό, στον αριθμό των σπερματοζωαρίων και στο ποσοστό κινητικότητας των σπερματοζωαρίων, αλλά προκάλεσε σημαντική μείωση στο επίπεδο των ανωμαλιών του σπέρματος. **ΣΥΜΠΕΡΑΣΜΑΤΑ** Η ανεκτικότητα του ανώτερου γαστρεντερικού σωλήνα και η αναπαραγωγική ασφάλεια αποδείχθηκαν στη χρόνια χρήση της σελεκοξίμπης σε άρρνες αλμπίνο αρουραίους, με βελτιωτικά αποτελέσματα του βασιλικού πολτού έναντι του οξειδωτικού και αποπτωτικού stress που προκαλείται από τη σελεκοξίμπη.

Λέξεις ευρητηρίου: Βασιλικός πολτός, Γαστρεντερικό, Όρχεις, Σελεκοξίμπη (celecoxib), Σπέρμα, Τεστοστερόνη

References

- PHILLIPS WJ, CURRIER BL. Analgesic pharmacology: II. Specific analgesics. *J Am Acad Orthop Surg* 2004, 12:221–223
- BINDU S, MAZUMDER S, BANDYOPADHYAY U. Non-steroidal anti-inflammatory drugs (NSAIDs) and organ damage: A current perspective. *Biochem Pharmacol* 2020, 180:114147
- GOLDSTEIN JL, CORREA P, ZHAO WW, BURR AM, HUBBARD RC, VERBURG KM ET AL. Reduced incidence of gastroduodenal ulcers with celecoxib, a novel cyclooxygenase-2 inhibitor, compared to naproxen in patients with arthritis. *Am J Gastroenterol* 2001, 96:1019–1027
- SHIN S. Safety of celecoxib versus traditional nonsteroidal anti-inflammatory drugs in older patients with arthritis. *J Pain Res* 2018, 11:3211–3219
- CHEN H, LUO L, LIU J, ZIRKIN BR. Cyclooxygenases in rat Leydig cells: Effects of luteinizing hormone and aging. *Endocrinology* 2007, 148:735–742
- ISHIKAWA T, HWANG K, LAZZARINO D, MORRIS PL. Sertoli cell expression of steroidogenic acute regulatory protein-related lipid transfer 1 and 5 domain-containing proteins and sterol regulatory element binding protein-1 are interleukin-1 β regulated by activation of c-Jun N-terminal kinase and cyclooxygenase-2 and cytokine induction. *Endocrinology* 2005, 146:5100–5111
- ISHIKAWA T, MORRIS PL. A multistep kinase-based Sertoli cell autocrine-amplifying loop regulates prostaglandins, their receptors, and cytokines. *Endocrinology* 2006, 147:1706–1716
- JAHANPOUR NS, JAHANPOUR F. The effect of non-steroidal drug in male rats' fertility. An experimental design. *Int J Fertil* 2013, 1:58
- JAHANPOUR N. Infertility in couple and celecoxib usage. An experimental design. *Int J Reprod Biomed* 2014, 12:105
- WINNALL WR, MUIR JA, LIEW S, HIRST JJ, MEACHEM SJ, HEDGER MP. Effects of chronic celecoxib on testicular function in normal and lipopolysaccharide-treated rats. *Int J Androl* 2009, 32:542–555
- WINNALL WR, ALI U, O'BRYAN MK, HIRST JJ, WHILEY PAF, MUIR JA ET AL. Constitutive expression of prostaglandin-endoperoxide synthase 2 by somatic and spermatogenic cells is responsible for prostaglandin E2 production in the adult rat testis. *Biol Reprod* 2007, 76:759–768
- CHATZIMELETIOU K, GALANIS N, KARAGIANNIDIS A, SIOGA A, PADOS G, GOULIS D ET AL. Fertility potential in a man with ankylosing spondylitis as revealed by semen analysis by light, electron and fluorescence microscopy. *SAGE Open Med Case Rep* 2018, 6:2050313X18759898
- CHATZIMELETIOU K, FLEVA A, SIOGA A, GEORGIU I, NIKOLOPOULOS TT, MARKOPOULOU M ET AL. Effects of different drug therapies and COVID-19 mRNA vaccination on semen quality in a man with ankylosing spondylitis: A case report. *Medicina (Kaunas)* 2022, 58:173
- AHMAD S, CAMPOS MG, FRATINI F, ALTAYE SZ, LI J. New insights into the biological and pharmaceutical properties of royal jelly. *Int J Mol Sci* 2020, 21:382
- VAZHACHARICKAL PJ. A review on health benefits and biological action of honey, propolis and royal jelly. *J Med Plants Stud* 2021, 9:1–3
- KOÇKAYA EA, SELMANOĞLU G, KISMET K, AKAY MT. Pathological and biochemical effects of therapeutic and supratherapeutic doses of celecoxib in Wistar albino male rats. *Drug Chem Toxicol* 2010, 33:410–414
- MOSTAFA RE, EL-MARASY SA, JALEEL GAA, BAKEER RM. Protective effect of royal jelly against diclofenac-induced hepato-renal damage and gastrointestinal ulcerations in rats. *Heliyon* 2020, 6:e03330
- SZABO S, HOLLANDER D. Pathways of gastrointestinal protection and repair: Mechanisms of action of sucralfate. *Am J Med* 1989, 86:23–31
- EL-NAGGAR DA, EL-ZALABANY LMA, SHAHIN DA, ATTIA AM, ELMOSALLAMY SA. Testicular toxicity of chloroxylenol in rats: Biochemical, pathological and flow cytometric study. *J Exp Pharmacol* 2022, 13:213–220
- ELEAWA SM, ALKHATEEB MA, ALHASHEM FH, BIN-JALIAH I, SAKR HF, ELREFAEY HM ET AL. Resveratrol reverses cadmium chloride-induced testicular damage and subfertility by downregulating p53 and Bax and upregulating gonadotropins and *Bcl-2* gene expression. *J Reprod Dev* 2014, 60:115–127
- NASERI MH, MAHDAVI M, DAVOODI J, TACKALLOU SH, GOUDARZVAND M, NEISHABOURI SH. Up regulation of Bax and down regulation of *Bcl2* during 3-NC mediated apoptosis in human cancer cells. *Cancer Cell Int* 2015, 15:55
- GRANOFF AB, ABRAHAM GE. Peripheral and adrenal venous levels of steroids in a patient with virilizing adrenal adenoma. *Obstet Gynecol* 1979, 53:111–115
- WANG M, UTELL MJ, SCHNEIDER A, ZAREBA W, FRAMPTON MW, OAKES D ET AL. Does total antioxidant capacity modify adverse cardiac responses associated with ambient ultrafine, accumulation mode, and fine particles in patients undergoing cardiac rehabilitation? *Environ Res* 2016, 149:15–22
- DIIVA S, MADHURI D, LAKSHMAN M, REDDY AG. Epididymal semen analysis in testicular toxicity of doxorubicin in male albino Wistar rats and its amelioration with quercetin. *J Pharm Innov* 2018, 7:555–558
- SCHÖNTHAL AH, CHEN TC, HOFMAN FM, LOUIE SG, PETASIS NA. Celecoxib analogs that lack COX-2 inhibitory function: Pre-clinical development of novel anticancer drugs. *Expert Opin Investig Drugs* 2008, 17:197–208
- HEGAZI MM, HELMY HM, SALIM EI. Celecoxib therapy attenuates oxidative stress during chemically induced rat colon carcinogenesis. *Egypt J Exp Biol (Zoo)* 2017, 13:177–185
- SHAO D, KAN M, QIAO P, PAN Y, WANG Z, XIAO X ET AL. Celecoxib induces apoptosis via a mitochondria-dependent pathway in the H22 mouse hepatoma cell line. *Mol Med Rep* 2014, 10:2093–2098
- GARCÍA-RAYADO G, NAVARRO M, LANAS A. NSAID induced gastrointestinal damage and designing GI-sparing NSAIDs. *Expert Rev Clin Pharmacol* 2018, 11:1031–1043
- MOHALE DS, TRIPATHI AS, WAHANE JB, CHANDEWAR AV. A pharmacological review on cyclooxygenase enzyme. *Indian J Pharm*

- Pharmacol* 2014, 1:46–58
30. GUDIS K, SAKAMOTO C. The role of cyclooxygenase in gastric mucosal protection. *Dig Dis Sci* 2005, 50(Suppl 1):S16–S23
 31. FAKI Y, ER A. Different chemical structures and physiological/pathological roles of cyclooxygenases. *Rambam Maimonides Med J* 2021, 12:e0003
 32. HALTER F, TARNAWSKI AS, SCHMASSMANN A, PESKAR BM. Cyclooxygenase 2-implications on maintenance of gastric mucosal integrity and ulcer healing: Controversial issues and perspectives. *Gut* 2001, 49:443–453
 33. PESKAR BM. Role of cyclooxygenase isoforms in gastric mucosal defense and ulcer healing. *Inflammopharmacology* 2005, 13:15–26
 34. WALLACE JL, DEVCHAND PR. Emerging roles for cyclooxygenase-2 in gastrointestinal mucosal defense. *Br J Pharmacol* 2005, 145:275–282
 35. GRETZER B, MARICIC N, RESPONDEK M, SCHULIGOI R, PESKAR BM. Effects of specific inhibition of cyclo-oxygenase-1 and cyclo-oxygenase-2 in the rat stomach with normal mucosa and after acid challenge. *Br J Pharmacol* 2001, 132:1565–1573
 36. WALLACE JL, MCKNIGHT W, REUTER BK, VERGNOLLE N. NSAID-induced gastric damage in rats: Requirement for inhibition of both cyclooxygenase 1 and 2. *Gastroenterology* 2000, 119:706–714
 37. TANAKA A, ARAKI H, KOMOIKE Y, HASE S, TAKEUCHI K. Inhibition of both COX-1 and COX-2 is required for development of gastric damage in response to nonsteroidal antiinflammatory drugs. *J Physiol Paris* 2001, 95:21–27
 38. LANGENBACH R, MORHAM SG, TIANO HF, LOFTIN CD, GHANAYEM BI, CHULADA PC ET AL. Prostaglandin synthase 1 gene disruption in mice reduces arachidonic acid-induced inflammation and indomethacin-induced gastric ulceration. *Cell* 1995, 83:483–492
 39. MORHAM SG, LANGENBACH R, LOFTIN CD, TIANO HF, VOULOU-MANOS N, JENNETTE JC ET AL. Prostaglandin synthase 2 gene disruption causes severe renal pathology in the mouse. *Cell* 1995, 83:473–482
 40. PALMER RH. COX-2 selectivity and lack of gastrointestinal toxicity: True, true and unrelated? *Gastroenterology* 2000, 118:640
 41. SOMASUNDARAM S, SIGTHORSSON G, SIMPSON RJ, WATTS J, JACOB M, TAVARES IA ET AL. Uncoupling of intestinal mitochondrial oxidative phosphorylation and inhibition of cyclooxygenase are required for the development of NSAID-enteropathy in the rat. *Aliment Pharmacol Ther* 2000, 14:639–650
 42. WALLACE JL, TIGLEY AW. New insights into prostaglandins and mucosal defence. *Aliment Pharmacol Ther* 1995, 9:227–235
 43. LIANG TY, DENG RM, LI X, XU X, CHEN G. The role of nitric oxide in peptic ulcer: A narrative review. *Med Gas Res* 2021, 11:42–45
 44. SENBEL AM, ABDELMONEIM L, OMAR AG. Celecoxib modulates nitric oxide and reactive oxygen species in kidney ischemia/reperfusion injury and rat aorta model of hypoxia/reoxygenation. *Vascul Pharmacol* 2014, 62:24–31
 45. EHRLICH K, SICKING C, RESPONDEK M, PESKAR BM. Interaction of cyclooxygenase isoenzymes, nitric oxide, and afferent neurons in gastric mucosal defense in rats. *J Pharmacol Exp Ther* 2004, 308:277–283
 46. SILVERSTEIN FE, FAICH G, GOLDSTEIN JL, SIMON LS, PINCUS T, WHELTON A ET AL. Gastrointestinal toxicity with celecoxib vs nonsteroidal anti-inflammatory drugs for osteoarthritis and rheumatoid arthritis: The CLASS study: A randomized controlled trial. Celecoxib Long-term Arthritis Safety Study. *JAMA* 2000, 284:1247–1255
 47. CRYER B. The role of cyclooxygenase selective inhibitors in the gastrointestinal tract. *Curr Gastroenterol Rep* 2003, 5:453–458
 48. CORUZZI G, VENTURI N, SPAGGIARI S. Gastrointestinal safety of novel nonsteroidal antiinflammatory drugs: Selective COX-2 inhibitors and beyond. *Acta Biomed* 2007, 78:96–110
 49. GOLDSTEIN JL, SILVERSTEIN FE, AGRAWAL NM, HUBBARD RC, KAISER J, MAURATH CJ ET AL. Reduced risk of upper gastrointestinal ulcer complications with celecoxib, a novel COX-2 inhibitor. *Am J Gastroenterol* 2000, 95:1681–1690
 50. CHAN FKL, CHING JYL, TSE YK, LAM K, WONG GLH, NG SC ET AL. Gastrointestinal safety of celecoxib versus naproxen in patients with cardiothrombotic diseases and arthritis after upper gastrointestinal bleeding (CONCERN): An industry-independent, double-blind, double-dummy, randomised trial. *Lancet* 2017, 389:2375–2382
 51. FENG GS, MA JL, WONG BCY, ZHANG L, LIU WD, PAN KF ET AL. Celecoxib-related gastroduodenal ulcer and cardiovascular events in a randomized trial for gastric cancer prevention. *World J Gastroenterol* 2008, 14:4535–4539
 52. GUO Q, LI Q, WANG J, LIU M, WANG Y, CHEN Z ET AL. A comprehensive evaluation of clinical efficacy and safety of celecoxib in combination with chemotherapy in metastatic or post-operative recurrent gastric cancer patients: A preliminary, three-center, clinical trial study. *Medicine (Baltimore)* 2019, 98:e16234
 53. MALLEEN SR, ESSEX MN, ZHANG R. Gastrointestinal tolerability of NSAIDs in elderly patients: A pooled analysis of 21 randomized clinical trials with celecoxib and nonselective NSAIDs. *Curr Med Res Opin* 2011, 27:1359–1366
 54. ABBAS B, ABBAS S, SATTAR M, RAHIM S, AMIR S. Comparison of efficacy of celecoxib (selective non-steroidal anti-inflammatory drug) versus naproxen (non-selective non-steroidal anti-inflammatory drug) for post endodontic pain. *Pak Armed Forces Med J* 2020, 31:1133–1137
 55. DURAN Y, KARABOĞA I, POLAT FR, POLAT E, ERBOĞA ZF, OVALI MA ET AL. Royal jelly attenuates gastric mucosal injury in a rat ethanol-induced gastric injury model. *Mol Biol Rep* 2020, 47:8867–8879
 56. ADEGBOYEGA PA, OLOLADE O. Immunohistochemical expression of cyclooxygenase-2 in normal kidneys. *Appl Immunohistochem Mol Morphol* 2004, 12:71–74
 57. BOTTING RM. Cyclooxygenase: Past, present and future. A tribute to John R. Vane (1927–2004). *J Therm Biol* 2006, 31:208–219
 58. MOHAN AR, BENNETT PR. Reproduction: Role of COX-2 and its inhibition. In: Pairet M, Ryn J (eds) *COX-2 inhibitors*. Birkhäuser Basel, Basel, 2004:213–225
 59. NEERAJA S, SREENATH AS, REDDY PRK, REDDANNA P. Expression of cyclooxygenase-2 in rat testis. *Reprod Biomed Online* 2003, 6:302–309

60. FRUNGIERI MB, GONZALEZ-CALVAR SI, MATZKIN ME, MAYERHOFER A, CALANDRA RS. Sources and functions of prostaglandins in the testis: Evidence for their relevance in male (in)fertility. *Anim Reprod* 2007, 4:63–69
61. PERROTTA I, SANTORO M, GUIDO C, AVENA P, TRIPEPI S, DE AMICIS F ET AL. Expression of cyclooxygenase-1 (COX-1) and COX-2 in human male gametes from normal patients, and those with varicocele and diabetes: A potential molecular marker for diagnosing male infertility disorders. *J Anat* 2012, 221:209–220
62. SCHELL C, FRUNGIERI MB, ALBRECHT M, GONZALEZ-CALVAR SI, KÖHN FM, CALANDRA RS ET AL. A prostaglandin D2 system in the human testis. *Fertil Steril* 2007, 88:233–236
63. BALAJIT, RAMANATHAN M, MENON VP. Localization of cyclooxygenase-2 in mice testis and assessment of its possible role through suppressing its expression using nimesulide: A preferential cyclooxygenase-2 inhibitor. *Prostaglandins Leukot Essent Fatty Acids* 2007, 76:341–348
64. FRUNGIERI MB, CALANDRA RS, MAYERHOFER A, MATZKIN ME. Cyclooxygenase and prostaglandins in somatic cell populations of the testis. *Reproduction* 2015, 149:R169–R180
65. HAMDULAY SS, WANG B, BIRDSEY GM, ALI F, DUMONT O, EVANS PC ET AL. Celecoxib activates PI-3K/Akt and mitochondrial redox signaling to enhance heme oxygenase-1-mediated anti-inflammatory activity in vascular endothelium. *Free Radic Biol Med* 2010, 48:1013–1023
66. KALE OE, OYESOLA TO, RAJI FS. Celecoxib, a cyclooxygenase-2 inhibitor, offers chemoprevention against reproductive and neurobehavioural abnormalities induced by atrazine in male Wistar rats. *Environ Toxicol Pharmacol* 2018, 58:84–97
67. MAZHARI S, RAZI M, MALEKINEJAD H, SADRKHANLOU R. Celecoxib and silymarin attenuated varicocele-induced damages at testicular and sperm levels; evidence for endocrine and antioxidant statuses. *Iran J Reprod Med* 2014, 12:117–119
68. HABIBI B, SADEGHIPOUR H, SEIFI B, MUGAHI MHN, SAEED TA. Protective effects of celecoxib on inflammatory cytokine levels and testis indices after induction of varicocele. *Physiol Pharmacol* 2014, 18:47–60
69. BALDERAS E, SÁNCHEZ-CÁRDENAS C, CHÁVEZ JC, DE LA VEGA BELTRÁN JL, GÓMEZ-LAGUNAS F, TREVIÑO CL ET AL. The anti-inflammatory drug celecoxib inhibits T-type Ca^{2+} currents in spermatogenic cells yet it elicits the acrosome reaction in mature sperm. *FEBS Lett* 2013, 587:2412–2419
70. YANG A, ZHOU M, ZHANG L, XIE G, CHEN H, LIU Z ET AL. Influence of royal jelly on the reproductive function of puberty male rats. *Food Chem Toxicol* 2012, 50:1834–1840
71. AMIRSHAHI T, NAJAFI G, NEJATI V. Protective effect of royal jelly on fertility and biochemical parameters in bleomycin-induced male rats. *Iran J Reprod Med* 2014, 2:209–216
72. ZAHMATKESH E, NAJAFI G, NEJATI V, HEIDARI R. Protective effect of royal jelly on the sperm parameters and testosterone level and lipid peroxidation in adult mice treated with oxymetholone. *Avicenna J Phytomed* 2014, 4:43–52
73. TAŞDOĞAN AM, PANCAR Z, ÖZDAL M, VURAL M, PANCAR S, BIRINCI YZ. The effect of short-term royal jelly supplement on testosterone levels in sedentary and healthy individuals. *Prog Nutr* 2020, 22:275–280
74. PEIVANDI S, SAVADKOUHI SK, ABBASI Z, ZAMANIYAN M, GORDANI N, MORADI S. Effect of royal jelly on sperm parameters and testosterone levels in infertile men. *J Mazandaran Univ Med Sci* 2022, 31:43–52
75. AL-SANAFI AE, MOHSSIN SA, ABDULLA SM. Effect of royal jelly on male infertility. *Thi-Qar Medical Journal* 2007, 1:1–2
76. GAWISH AM, EIFIKY S, THERASE M, ABDELRAOF A, KHALIL W, MOHAMED KA. Sperm abnormality toxicity due to cyclosporine A and the ameliorative effect of royal jelly in male rats. *J Basic Appl Zool* 2016, 76:60–73
77. MAHDIVAND N, NAJAFI G, NEJATI V, SHALIZAR-JALALI A, RAHMANI F. Royal jelly protects male rats from heat stress-induced reproductive failure. *Andrologia* 2019, 51:e13213
78. TREVISAN M, BROWNE R, RAM M, MUTI P, FREUDENHEIM J, CARSELLA AM ET AL. Correlates of markers of oxidative status in the general population. *Am J Epidemiol* 2001, 154:348–356
79. BERNABUCCI U, RONCHI B, LACETERA N, NARDONE A. Markers of oxidative status in plasma and erythrocytes of transition dairy cows during hot season. *J Dairy Sci* 2002, 85:2173–2179
80. RAO PS, KALVA S, YERRAMILI A, MAMIDI S. Free radicals and tissue damage: Role of antioxidants. *Free Radic Antioxid* 2011, 1:2–7
81. DUPRÉ-CROCHET S, ERARD M, NÜSSE O. ROS production in phagocytes: Why, when, and where? *J Leukoc Biol* 2013, 94:657–670
82. VALKO M, LEIBFRITZ D, MONCOL J, CRONIN MTD, MAZUR M, TELSNER J. Free radicals and antioxidants in normal physiological functions and human disease. *Int J Biochem Cell Biol* 2007, 39:44–84
83. PHAM-HUY LA, HE H, PHAM-HUY C. Free radicals, antioxidants in disease and health. *Int J Biomed Sci* 2008, 4:89–96
84. ANAND DAVID AV, ARULMOLI R, PARASURAMAN S. Overviews of biological importance of quercetin: A bioactive flavonoid. *Pharmacogn Rev* 2016, 10:84–89
85. ĎURAČKOVÁ Z. Some current insights into oxidative stress. *Physiol Res* 2010, 59:459–469
86. REITER RJ, PAREDES SD, MANCHESTER LC, TAN DX. Reducing oxidative/nitrosative stress: A newly-discovered genre for melatonin. *Crit Rev Biochem Mol Biol* 2009, 44:175–200
87. ZIECH D, FRANCO R, GEORGAKILAS AG, GEORGAKILA S, MALAMOU-MITSIS V, SCHONEVELD O ET AL. The role of reactive oxygen species and oxidative stress in environmental carcinogenesis and biomarker development. *Chem Biol Interact* 2010, 188:334–339
88. MIROŃCZUK-CHODAKOWSKA I, WITKOWSKA AM, ZUJKO ME. Endogenous non-enzymatic antioxidants in the human body. *Adv Med Sci* 2018, 63:68–78
89. CIMEN MYB, CIMEN ÖB, ESKANDARI G, SAHIN G, ERDOĞAN C, ATIK U. *In vivo* effects of meloxicam, celecoxib, and ibuprofen on free radical metabolism in human erythrocytes. *Drug Chem Toxicol* 2003, 26:169–176
90. OZGOCMEN S, ARDICOGLU O, ERDOGAN H, FADILIOGLU E, GUDUL H. *In vivo* effect of celecoxib and tenoxicam on oxidant/antioxidant status of patients with knee osteoarthritis. *Ann Clin Lab Sci* 2005, 35:137–143
91. SOZER S, DINIZ G, LERMIOGLU F. Effects of celecoxib in young rats: Histopathological changes in tissues and alterations of

- oxidative stress/antioxidant defense system. *Arch Pharm Res* 2011, 34:253-259
92. MELEKH B, ILKIV I, LOZYNSKYI A, SKLYAROV A. Antioxidant enzyme activity and lipid peroxidation in rat liver exposed to celecoxib and lansoprazole under epinephrine-induced stress. *J Appl Pharm Sci* 2017, 7:94-99
 93. HAMZA RZ, AL-EISA RA, EL-SHENAWY NS. Possible ameliorative effects of the royal jelly on hepatotoxicity and oxidative stress induced by molybdenum nanoparticles and/or cadmium chloride in male rats. *Biology (Basel)* 2022, 11:450
 94. COTTER TG, RINELLA M. Nonalcoholic fatty liver disease 2020: The state of the disease. *Gastroenterology* 2020, 158:1851-1864
 95. YOU MM, LIU YC, CHEN YF, PAN YM, MIAO ZN, SHI YZ ET AL. Royal jelly attenuates nonalcoholic fatty liver disease by inhibiting oxidative stress and regulating the expression of circadian genes in ovariectomized rats. *J Food Biochem* 2020, 44:e13138
 96. KARIMI E, KHORVASH F, ARAB A, SEPIDARKISH M, SAADATNIA M, AMANI R. The effects of royal jelly supplementation on oxidative stress, inflammatory mediators, mental health, cognitive function, quality of life, and clinical outcomes of patients with ischemic stroke: Study protocol for a randomized controlled trial. *BMC Nutr* 2023, 9:32
 97. KOHNO K, OKAMOTO I, SANO O, ARAI N, IWAKI K, IKEDA M ET AL. Royal jelly inhibits the production of proinflammatory cytokines by activated macrophages. *Biosci Biotechnol Biochem* 2004, 68:138-145
 98. ASLAN Z, AKSOY L. Anti-inflammatory effects of royal jelly on ethylene glycol induced renal inflammation in rats. *Int Braz J Urol* 2015, 41:1008-10013
 99. HADI A, NAJAFGHOLIZADEH A, AYDENLU ES, SHAFIEI Z, PIRIVAND F, GOLPOUR S ET AL. Royal jelly is an effective and relatively safe alternative approach to blood lipid modulation: A meta-analysis. *J Funct Foods* 2018, 41:202-209
 100. MALEKI V, JAFARI-VAYGHAN H, SALEH-GHADIMI S, ADIBIAN M, KHEIROURI S, ALIZADEH M. Effects of royal jelly on metabolic variables in diabetes mellitus: A systematic review. *Complement Ther Med* 2019, 43:20-27
 101. KANBUR M, ERASLAN G, SILICI S, KARABACAK M. Effects of sodium fluoride exposure on some biochemical parameters in mice: Evaluation of the ameliorative effect of royal jelly applications on these parameters. *Food Chem Toxicol* 2009, 47:1184-1189
 102. ELMORE S. Apoptosis: A review of programmed cell death. *Toxicol Pathol* 2007, 35:495-516
 103. RATHORE S, DATTA G, KAUR I, MALHOTRA P, MOHMMED A. Disruption of cellular homeostasis induces organelle stress and triggers apoptosis-like cell-death pathways in malaria parasite. *Cell Death Dis* 2015, 6:e1803
 104. HUSSAR P. Apoptosis regulators *bcl-2* and caspase-3. *Encyclopedia* 2022, 2:1624-1636
 105. LOCKSHIN RA, ZAKERI Z. Programmed cell death and apoptosis: Origins of the theory. *Nat Rev Mol Cell Biol* 2001, 2:545-550
 106. BLAGOSKLONNY MV. Cell death beyond apoptosis. *Leukemia* 2000, 14:1502-1508
 107. FINK SL, COOKSON BT. Apoptosis, pyroptosis, and necrosis: Mechanistic description of dead and dying eukaryotic cells. *Infect Immun* 2005, 73:1907-1916
 108. XU X, LAI Y, HUA ZC. Apoptosis and apoptotic body: Disease message and therapeutic target potentials. *Biosci Rep* 2019, 39:BSR20180992
 109. D'ARCY MS. Cell death: A review of the major forms of apoptosis, necrosis and autophagy. *Cell Biol Int* 2019, 43:582-592
 110. CHI H, CHANGHY, SANGTK. Neuronal cell death mechanisms in major neurodegenerative diseases. *Int J Mol Sci* 2018, 19:3082
 111. CHOI SM, CHO YS, PARK G, LEE SK, CHUN KS. Celecoxib induces apoptosis through Akt inhibition in 5-fluorouracil-resistant gastric cancer cells. *Toxicol Res* 2021, 37:25-33
 112. MANNING BD, TOKER A. AKT/PKB signaling: Navigating the network. *Cell* 2017, 169:381-405
 113. GAO D, NYALALI AM, HOU Y, XU Y, ZHOU J, ZHAO W ET AL. 2,5-dimethyl celecoxib inhibits proliferation and cell cycle and induces apoptosis in glioblastoma by suppressing CIP2A/PP2A/Akt signaling axis. *J Mol Neurosci* 2021, 71:1703-1713
 114. KARADENIZ A, SIMSEK N, KARAKUS E, YILDIRIM S, KARA A, CAN I ET AL. Royal jelly modulates oxidative stress and apoptosis in liver and kidneys of rats treated with cisplatin. *Oxid Med Cell Longev* 2011, 2011:981793
 115. AZAD F, NEJATIV, SHALIZAR-JALALI A, NAJAFI G, RAHMANI F. Antioxidant and anti-apoptotic effects of royal jelly against nicotine-induced testicular injury in mice. *Environ Toxicol* 2019, 34:708-718
 116. MOHAMED HK, MOBASHER MA, EBIIYA RA, HASSEN MT, HAGAG HM, EL-SAYED R ET AL. Anti-inflammatory, anti-apoptotic, and antioxidant roles of honey, royal jelly, and propolis in suppressing nephrotoxicity induced by doxorubicin in male albino rats. *Antioxidants (Basel)* 2022, 11:1029
 117. TOHAMY HG, EL-NEWESHY MS, SOLIMAN MM, SAYED S, SHUKRY M, GHAMRY HI ET AL. Protective potential of royal jelly against hydroxyurea-induced hepatic injury in rats via antioxidant, anti-inflammatory, and anti-apoptosis properties. *PLoS One* 2022, 17:e0265261
 118. FAZILI N, SOHEILI ZS, MALEKZADEH-SHAFAROUZI S, SAMIEI S, ALIPOOR SD, MOSHTAGHI N ET AL. Decreases MMP-9 expression and induces apoptosis in human bladder cancer 5637 cells. *Journal of Cell and Molecular Research (JCMR)* 2021, 13:36-43
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